



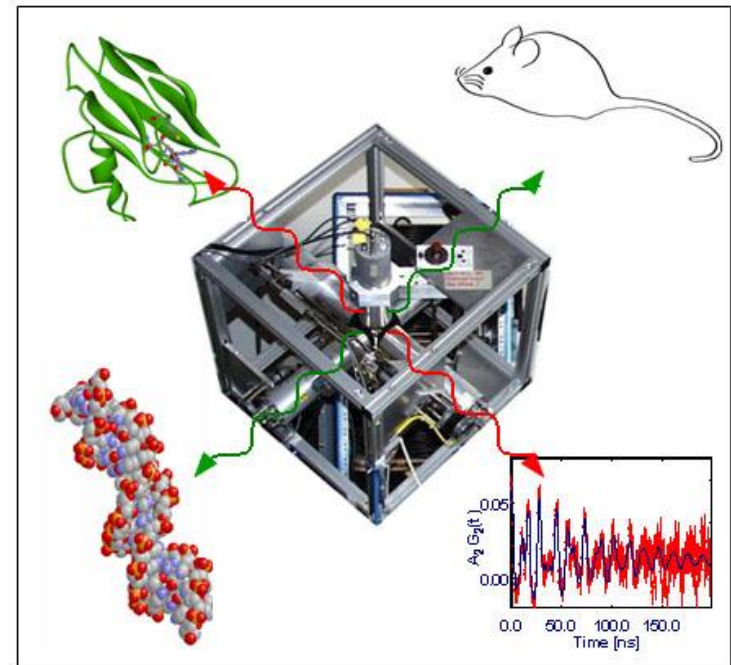
Biochemical applications of perturbed angular correlation of γ -rays (PAC) spectroscopy

Lars Hemmingsen (lhe@life.ku.dk), HFI/NQI 2010



Outline

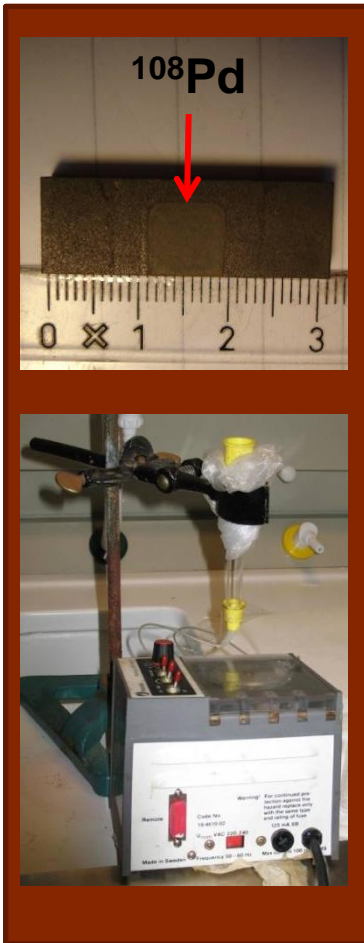
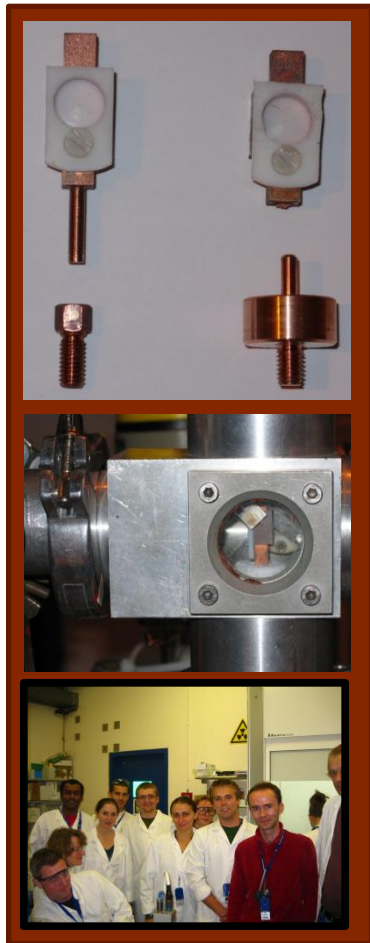
- Bio PAC introduction
- Selected examples:
 - *De novo* design of metal ion binding sites in proteins
 - Protein folding and misfolding
 - Catalysis – enzymes
 - Protein-protein interactions
 - *In vivo* experiments



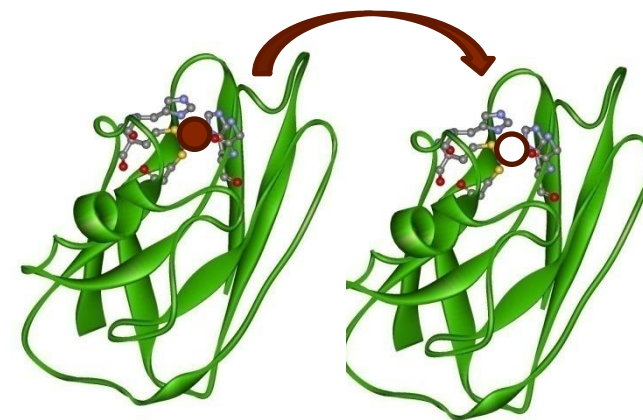
Bio PAC sample preparation

ISOLDE/CERN

Copenhagen



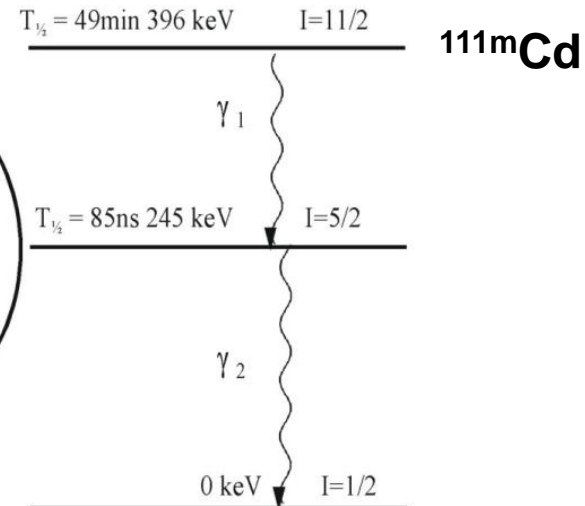
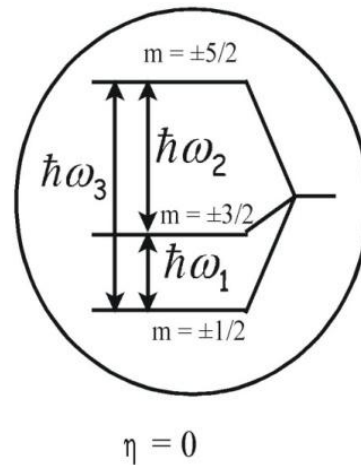
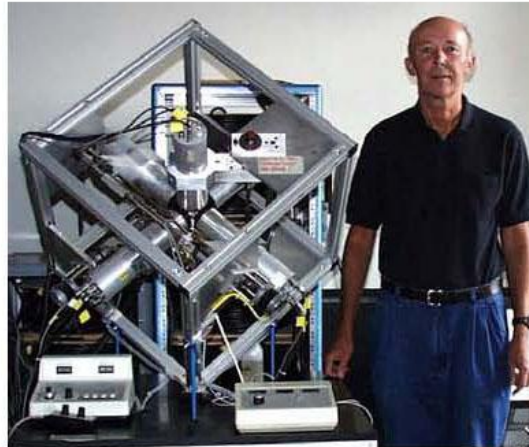
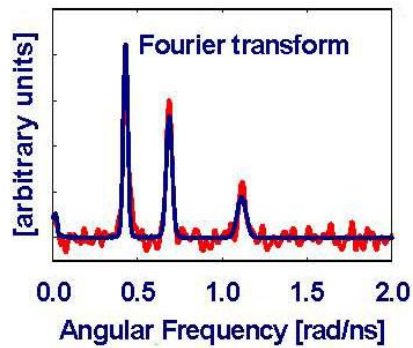
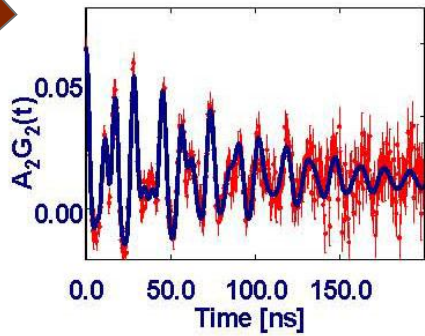
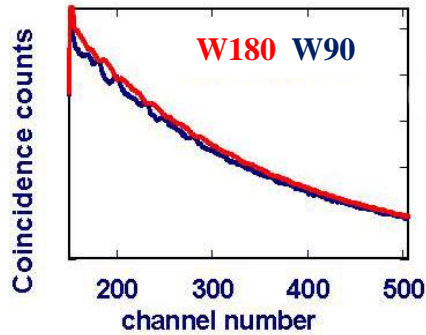
Metal ion replacement



Biomolecule in buffered solution
+ PAC probe (+carrier)
& immobilisation (sucrose, flash freeze, precipitation)

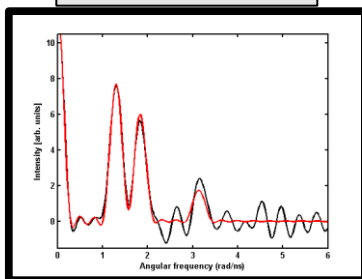
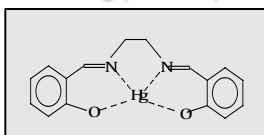


PAC in a nutshell

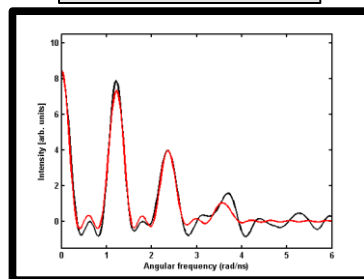
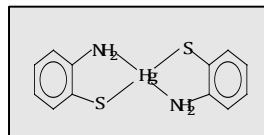


Interpretation of data: Model complexes and first principle calculations

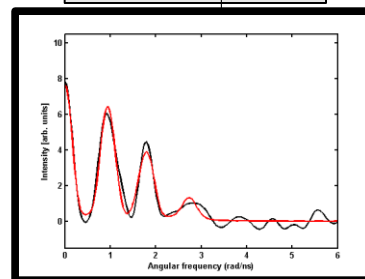
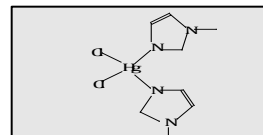
Hg(salen)



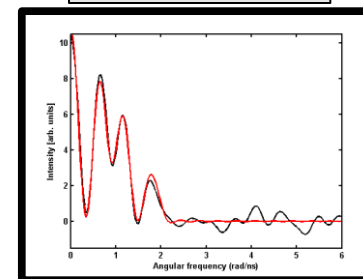
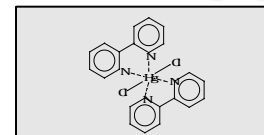
Hg(2-mercaptoanilin)₂



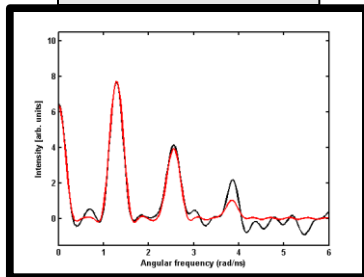
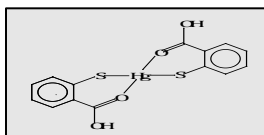
Hg(1-methylimidazole)₂Cl₂



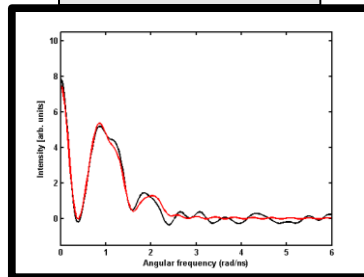
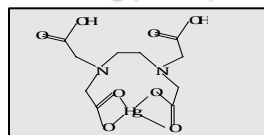
Hg(2,2'-bipy)₂Cl₂



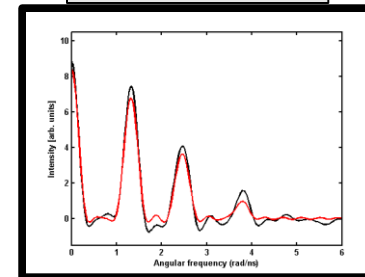
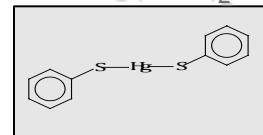
Hg(thiosalicylate)₂



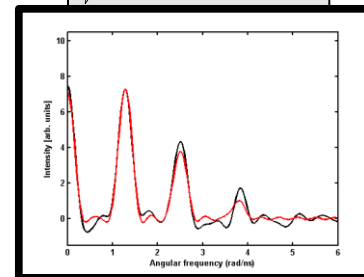
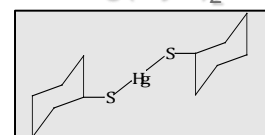
Hg(edta)



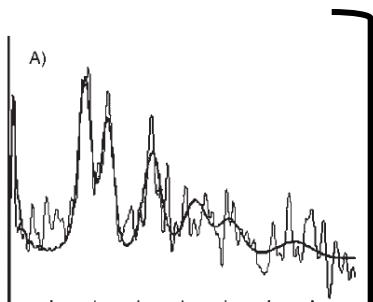
Hg(PhS)₂



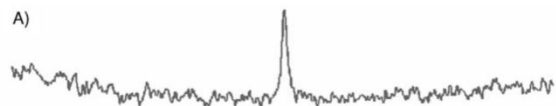
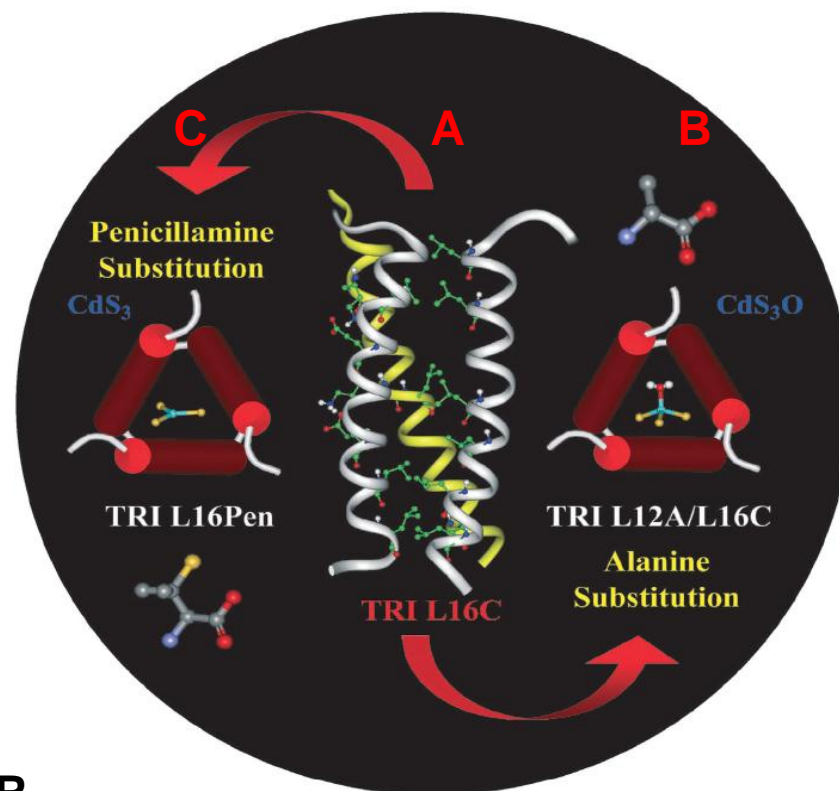
Hg(CyS)₂



De novo designed heavy metal ion binding proteins



^{111}mCd -PAC



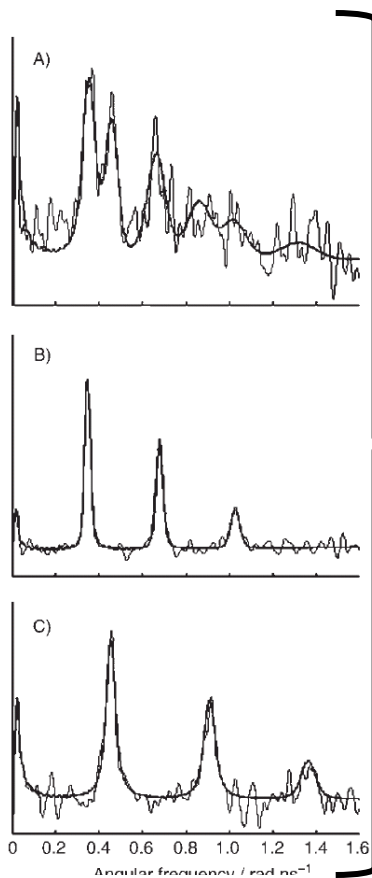
^{113}Cd -NMR

Chemical shift / ppm

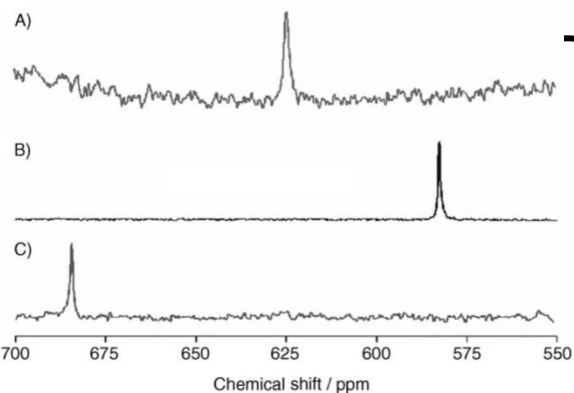
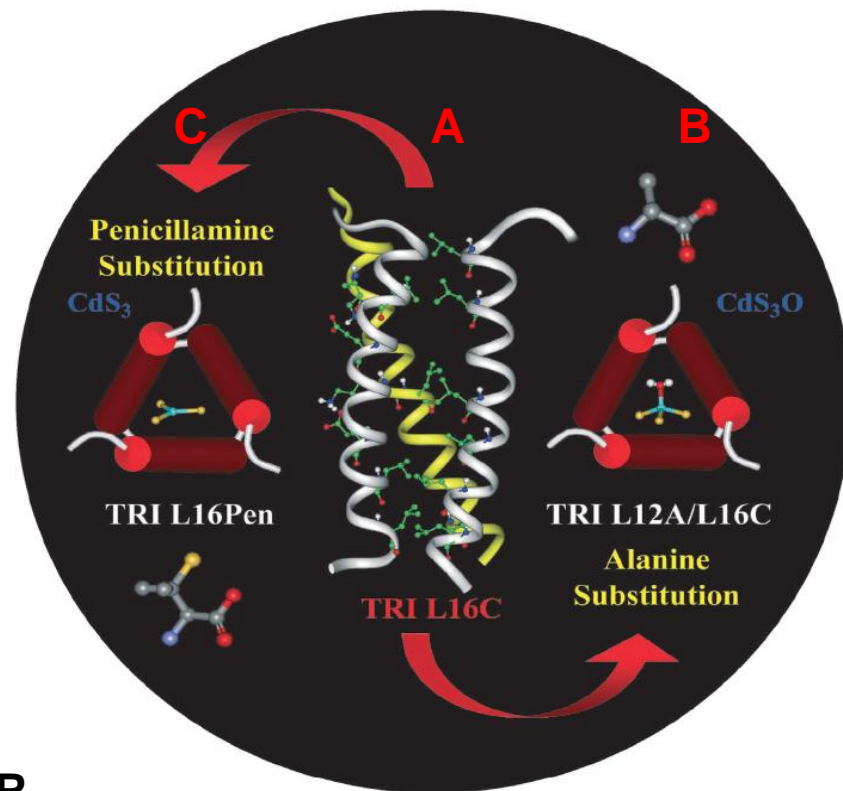
Matzapetakis *et al.* J. Am. Chem. Soc. **2002**, 124: 8042; Lee *et al.* Angew. Chem., **2006**, 45: 2864; Peacock *et al.* Proc. Nat. Acad. Sci. **2008**, 105: 16566



De novo designed heavy metal ion binding proteins



^{111m}Cd -PAC



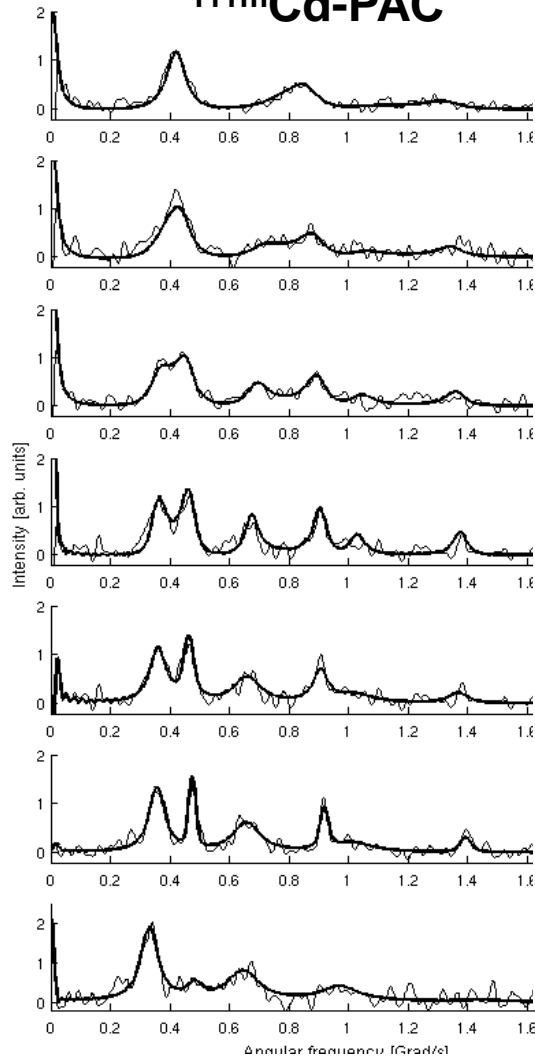
^{113}Cd -NMR

Matzapetakis *et al.* *J. Am. Chem. Soc.* **2002**, 124: 8042; Lee *et al.* *Angew. Chem.*, **2006**, 45: 2864; Peacock *et al.* *Proc. Nat. Acad. Sci.* **2008**, 105: 16566



De novo designed heavy metal Ion binding proteins: ns dynamics

^{111m}Cd-PAC



50 °C

35 °C

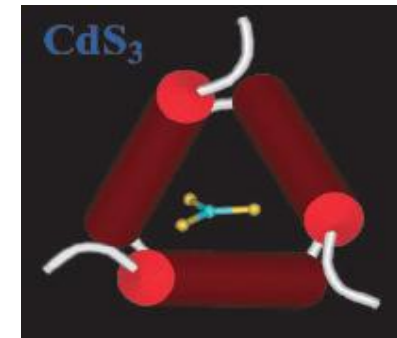
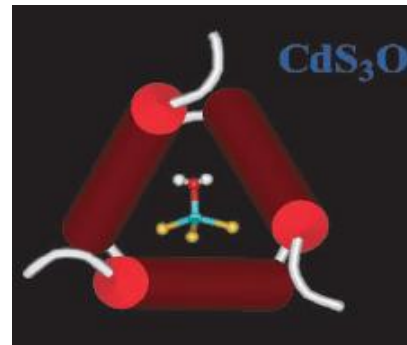
20 °C

1 °C

1 °C

-20 °C

-196 °C



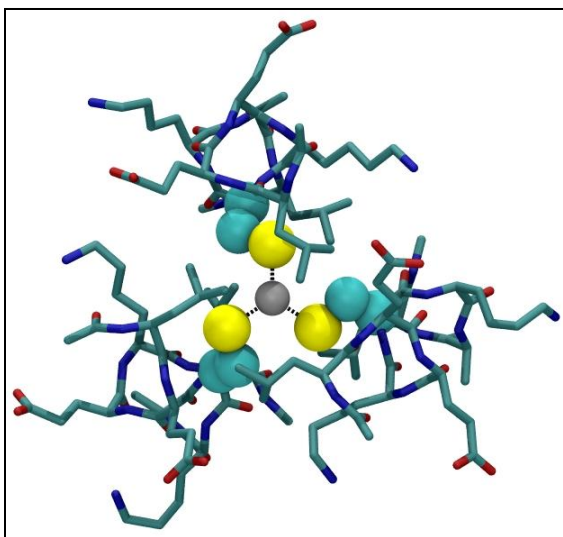
Temp [°C]	τ_1 [ns]	τ_{-1} [ns]
1	52	48
20	42	36
35	28	20
50	19	12

Stachura et al. Manuscript in preparation



Quantum mechanical structure optimization and property calculation

	ν_Q (MHz)	η	δ_{average} (ppm)	$\text{Cd-S}_{\text{average}}$ (Å)
41% CdS_3	466 465	0.16 0.20	647 624	2.48 2.49
59% CdS_3O	340 355	0.43 0.22		



Experimental data:

^{111}mCd -PAC, ^{113}Cd -NMR, EXAFS

Calculations:

ONIOM (Gaussian03)

PBE1PBE/6-31G(d), LanLDZ and PM3MM for structures;

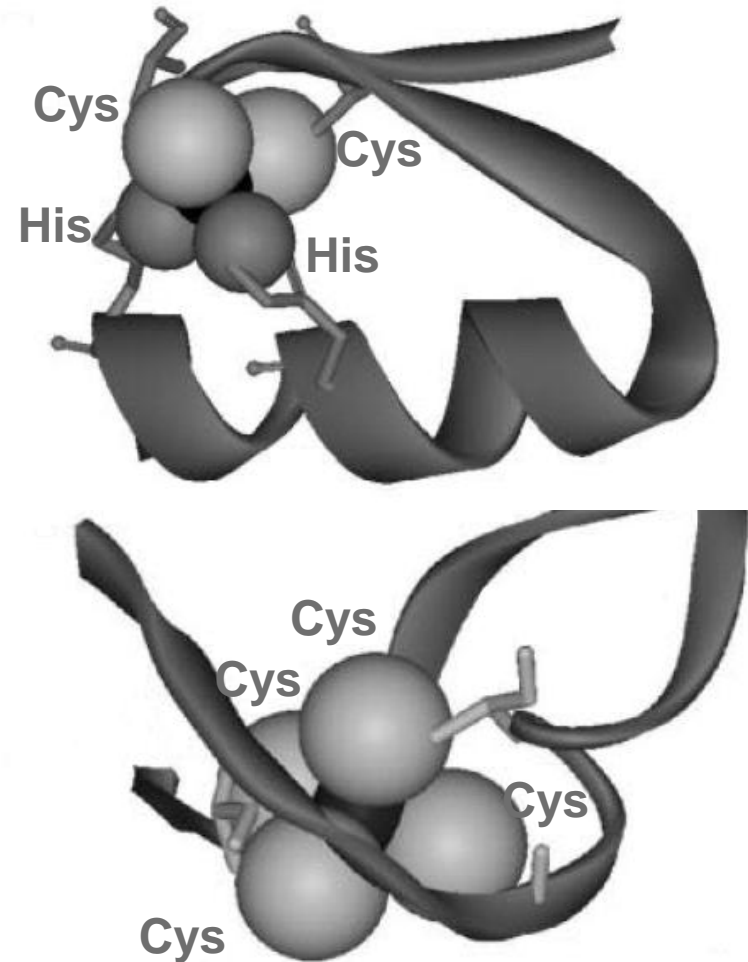
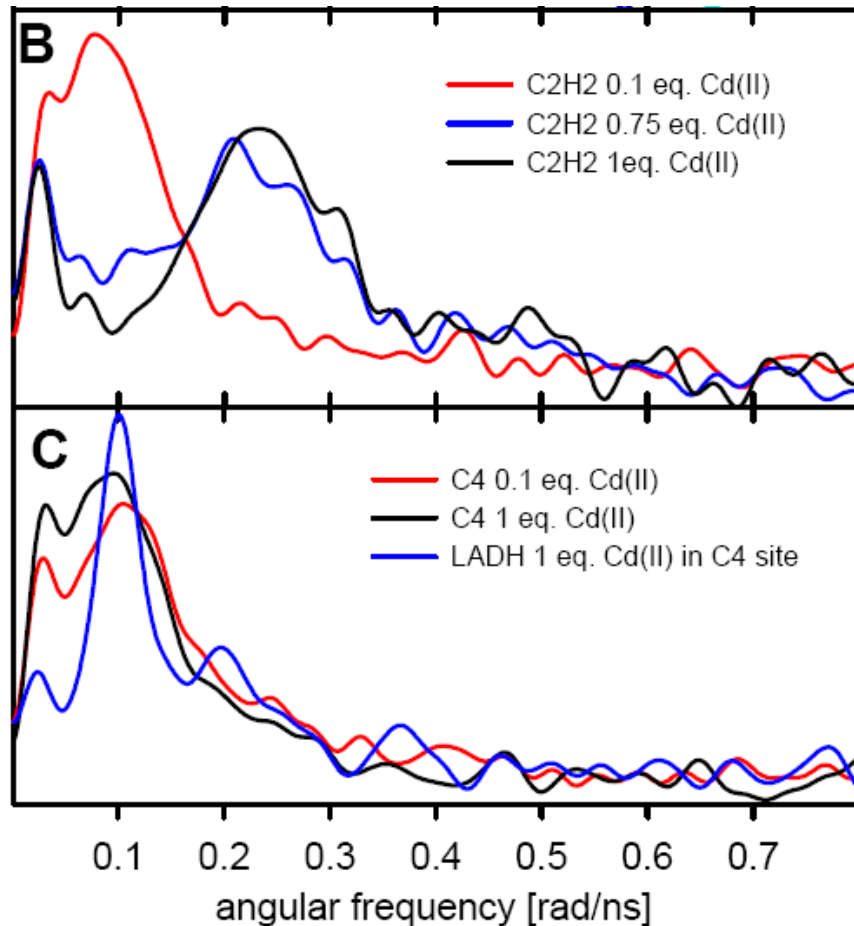
B3LYP/6-31G(d), Kellö & Sadlej for properties

Hemmingsen et al. J. Biol. Inorg. Chem. **2004**, 9, 591; Antony et al. J. Phys. Chem. **2000**, 104, 6047;
Hemmingsen and Ryde **1996**, J. Phys. Chem. 100, 4803, N.J. Christensen, Master thesis, **2005**, the Technical University of Denmark



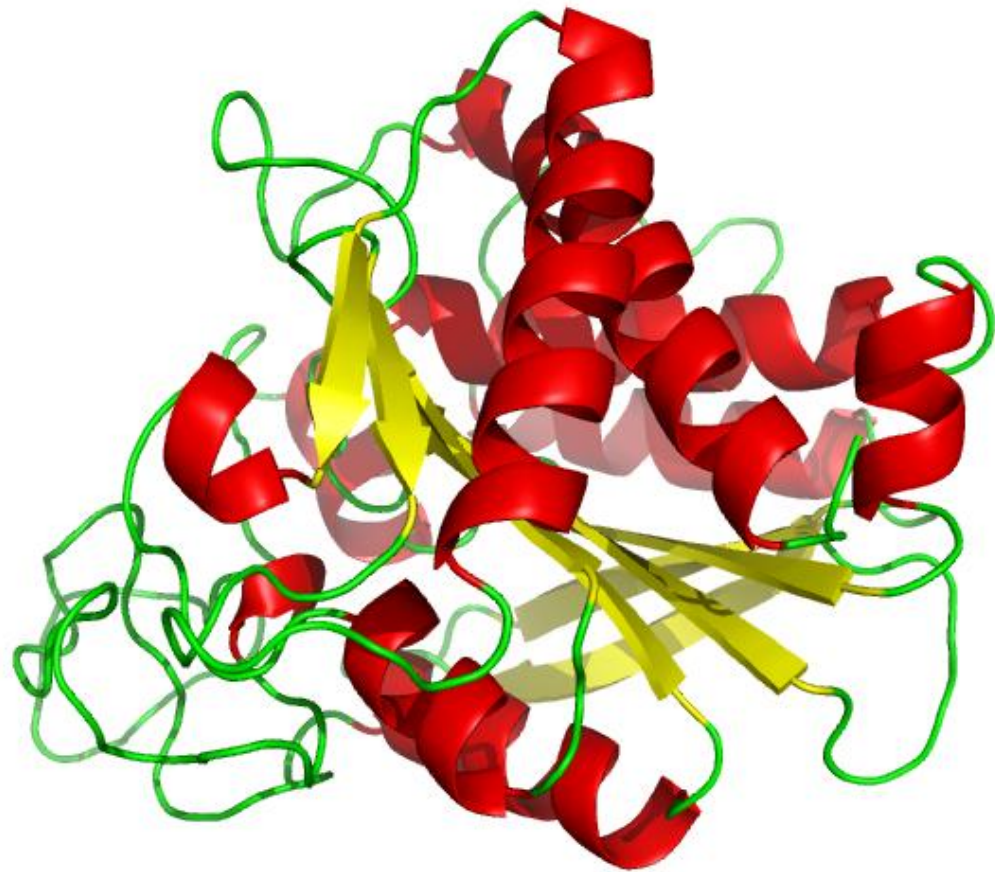
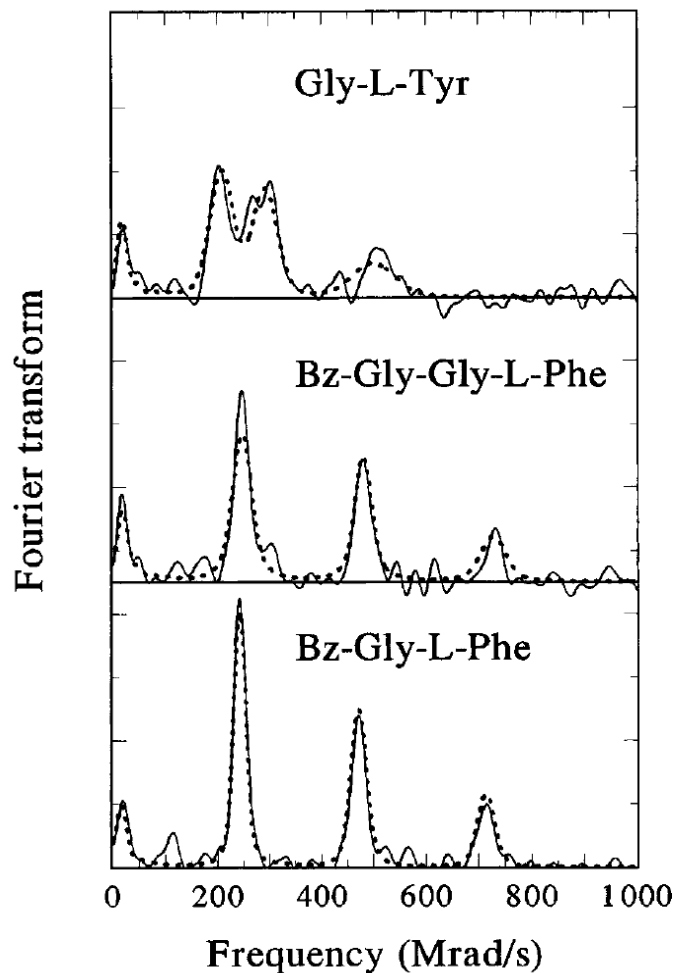
Metal ion controlled protein folding and misfolding

^{111m}Cd -PAC



Metal ion binding site structure during steady state catalysis (carboxypeptidase A)

^{111}mCd -PAC



Bauer et al Biochemistry 1997, 36, 11514;
[http://upload.wikimedia.org/wikipedia/commons/
5/5b/Carboxypeptidase_A.png](http://upload.wikimedia.org/wikipedia/commons/5/5b/Carboxypeptidase_A.png)



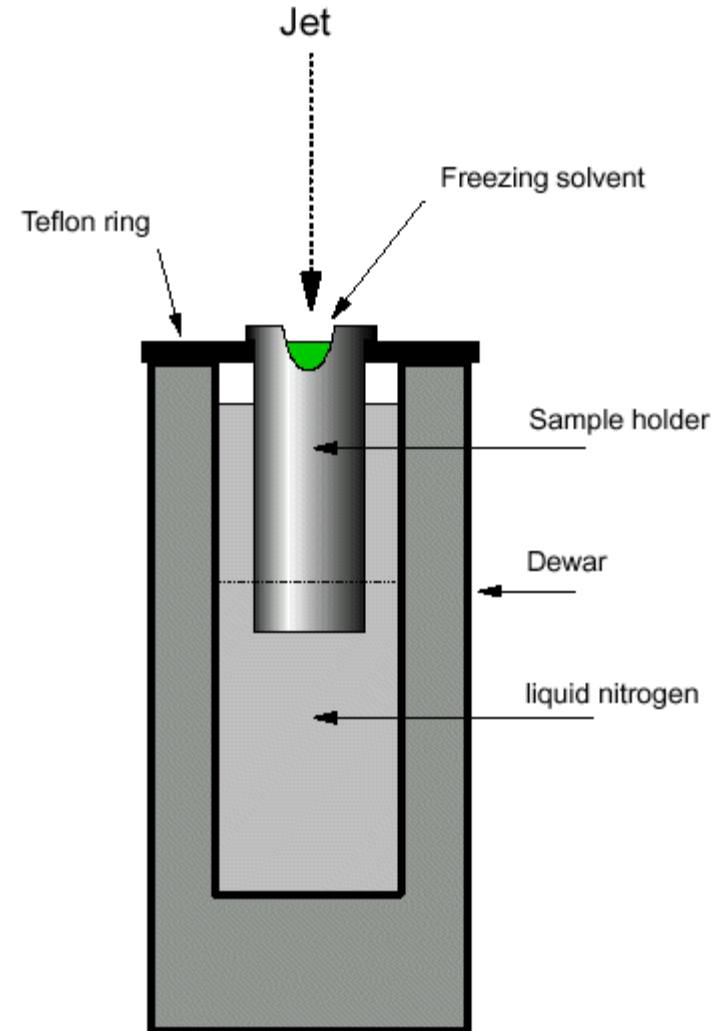
Freeze quench PAC spectroscopy:

Snap shots of structures evolving during enzyme catalyzed reactions

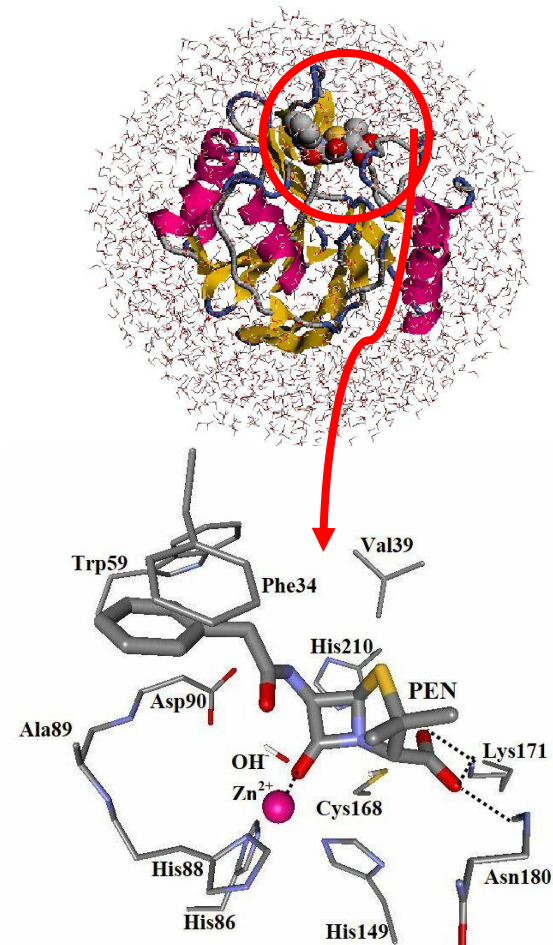
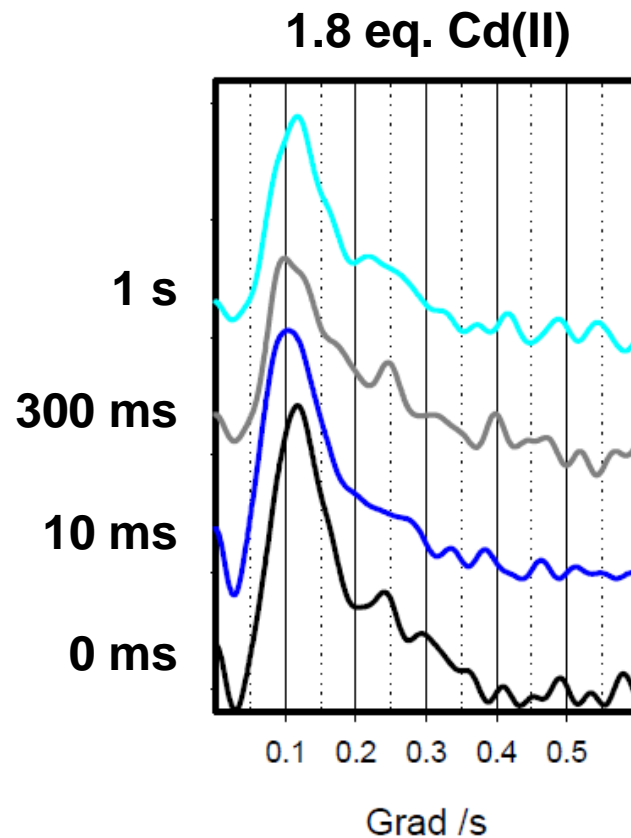


Enzyme

Substrate



Bacterial resistance to antibiotics – β -lactamases

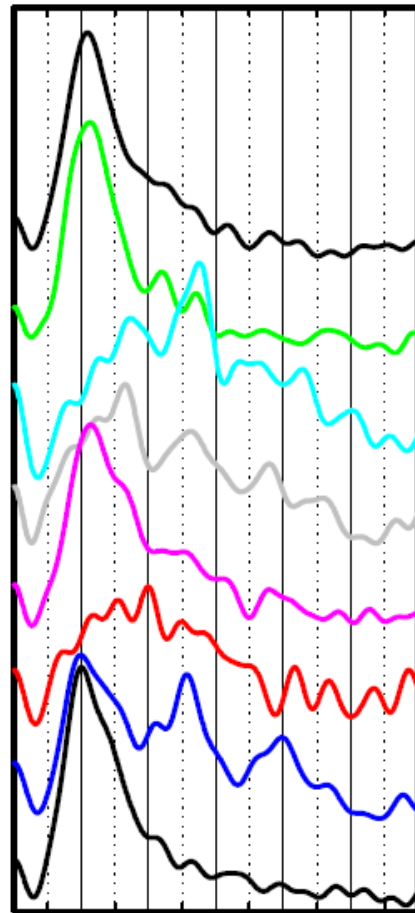


Heinz et al. in prep.; Olsen et al. *J.Phys. Chem. A*, **2002**, 106:1046; Olsen et al. *J.Phys. Chem. B*, **2003**, 107:2366; Olsen et al. *J.Phys. Chem. B*, **2004**, 108:17639



Bacterial resistance to antibiotics – β -lactamases

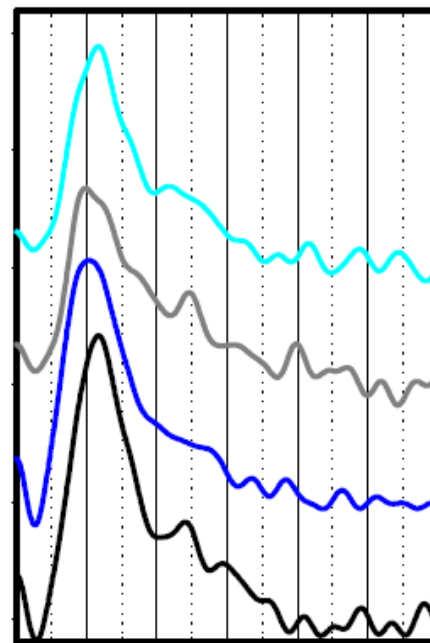
0.2 eq. Cd(II)



0.1 0.2 0.3 0.4 0.5

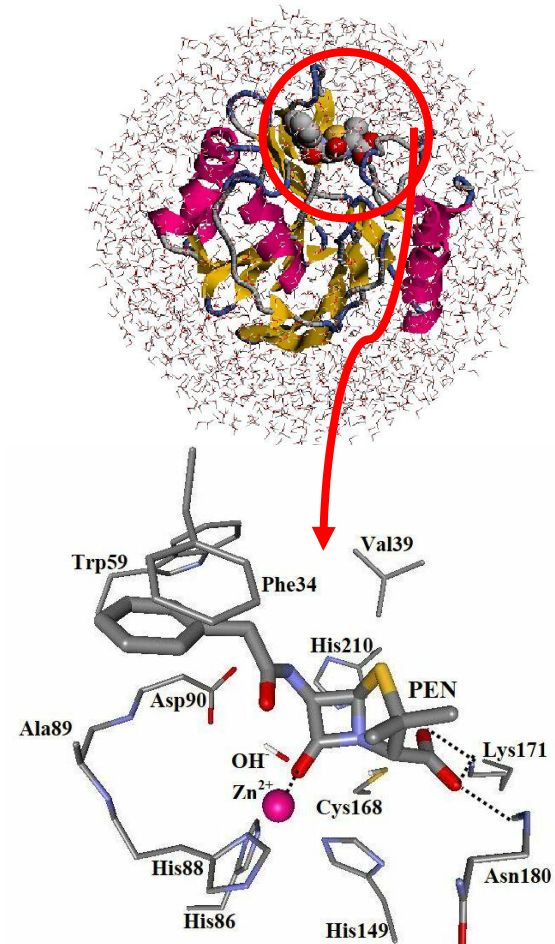
Grad / s

1.8 eq. Cd(II)



0.1 0.2 0.3 0.4 0.5

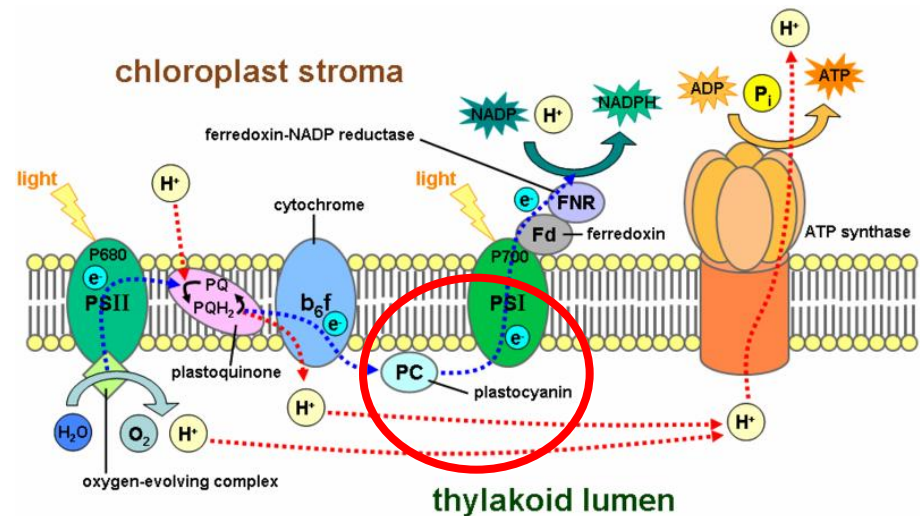
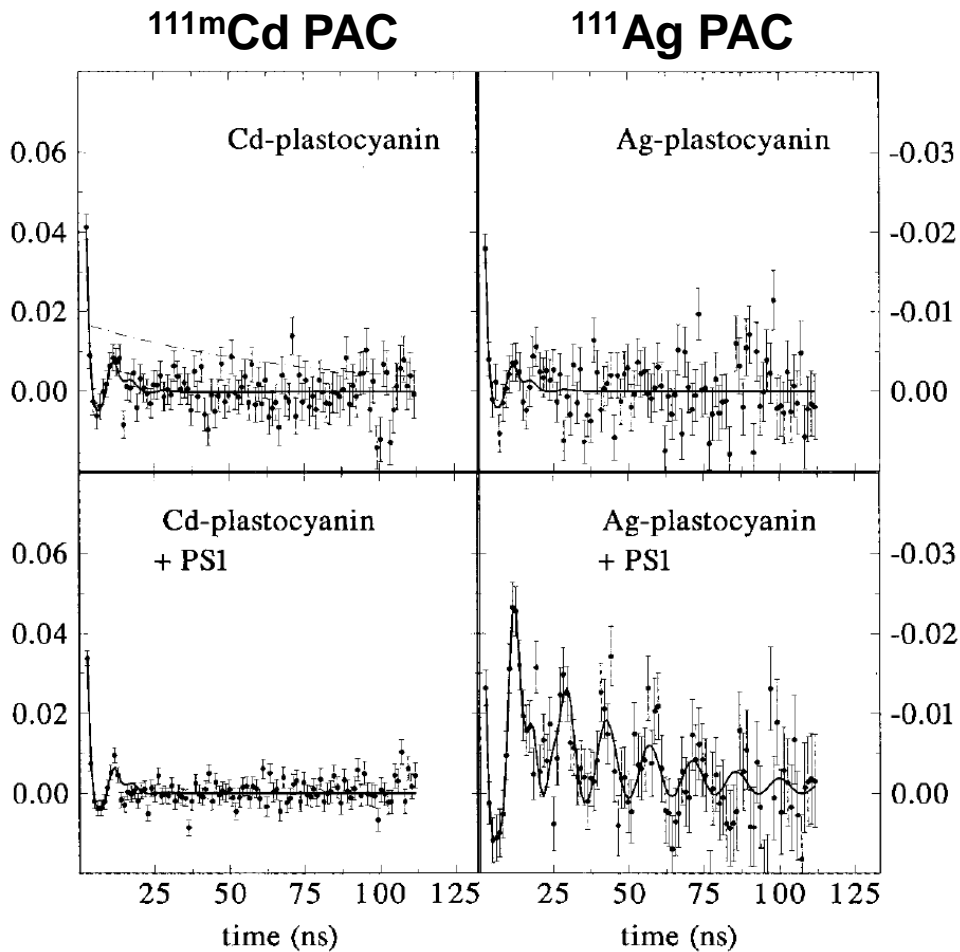
Grad / s



Heinz et al. in prep.; Olsen et al. *J.Phys. Chem. A*, **2002**, 106:1046; Olsen et al. *J.Phys. Chem. B*, **2003**, 107:2366; Olsen et al. *J.Phys. Chem. B*, **2004**, 108:17639



Protein-protein interactions: Plastocyanin binding to photosystem 1

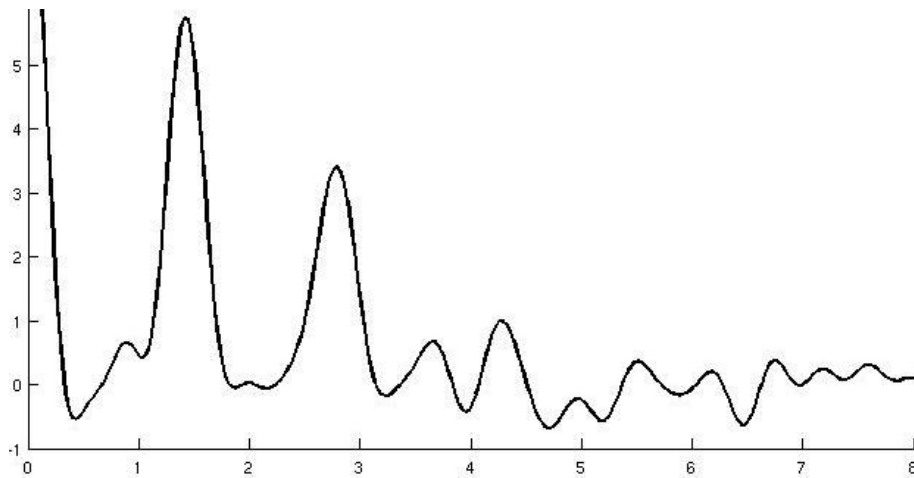


Danielsen et al. *Biochemistry*, 1999, 38:11531;
<http://en.wikipedia.org/wiki/Photosynthesis>

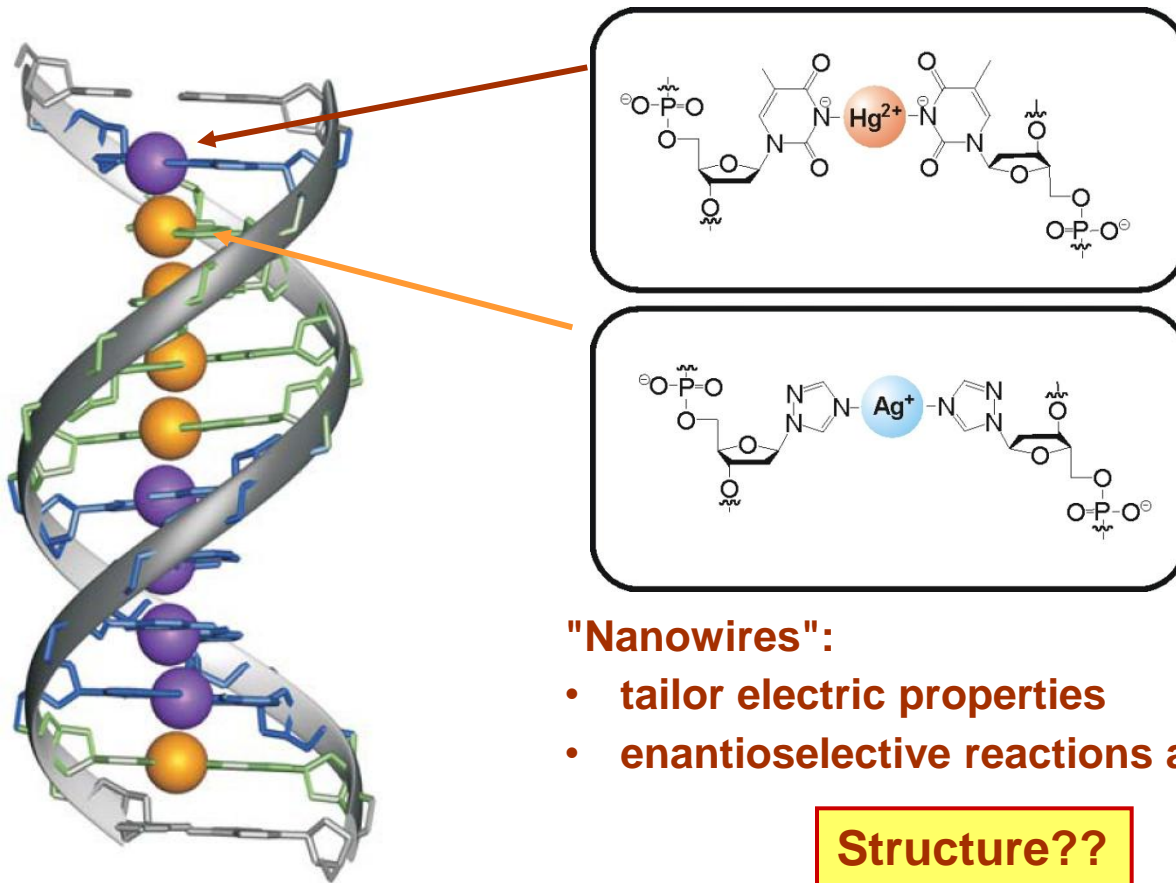


In vivo PAC experiments: Hg(II) binding to barley

^{199m}Hg -PAC



Molecular wires based on metal-modified nucleic acids



"Nanowires":

- tailor electric properties
- enantioselective reactions at the metal center

Structure??

Advantages and limitations of PAC-spectroscopy

Advantages:

- Characterisation of structure and dynamics at the PAC probe site (including rotational correlation times)
- High sensitivity to structural changes, e.g. during enzyme catalysis
- Small amount of PAC probe needed (in principle about 1 pmol)
- Different physical states (crystals, surfaces, solutions, *in vivo*...)
- Mechanically stable, allowing for stirring, flow, ...

Limitations:

- Suitable PAC isotopes do not exist for all elements
- PAC isotope must bind strongly to the molecule of interest
- Spectral parameters do not uniquely determine structure
- After effects can cause problems (in particular for EC)
- Production of PAC-isotopes

Hemmingsen et al. *Chem. Rev.*, **2004**, 104: 4027; Hemmingsen and Butz, in "Application of Physical Methods to Inorganic and Bioinorganic Chemistry" **2007**, Ed. R.A. Scott, Wiley



Acknowledgements



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- Universität Leipzig, Germany
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- Lunds Universitet, Sweden
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Ministry of Science
Technology and Innovation



DCSC



Danish Center for Scientific Computing

VILLUM KANN RASMUSSEN FONDEN
& VELUX FONDEN

